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胆管上皮细胞 TLR3 内源性活化在原发性胆汁性肝硬化中的作用

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摘要 目的:探讨胆管上皮细胞中 Toll 样受体 3 (TLR3) 内源性活化对原发性胆汁性肝硬化损伤的影响。**方法:**体外培养人肝内胆管上皮细胞 (HiBEC), 并通过冻融处理制备坏死 HiBEC 样本。将死亡样本与活样本混合培养 6 h, 以制备 TLR3 内源性活化样本, 将其标记为观察组; 以核酸酶处理坏死细胞及活细胞, 并混合培养 6 h, 以制备对照样本, 将其标记为对照组。检测 2 组细胞凋亡率、TLR3 及 β 干扰素 Toll 样受体结构域衔接蛋白 (TRIF) 表达水平、Caspase-3 活性。**结果:**观察组凋亡率明显高于对照组, TLR3 及 TRIF 相对表达量明显高于对照组, 细胞 Caspase-3 平均表达量明显高于对照组, 上述差异均有统计学意义 ($P < 0.05$)。**结论:**TLR3 的内源性活化可导致 HiBEC 凋亡, 在原发性胆汁性肝硬化的发生及发展中可能有一定促进作用。

关键词: Toll 样受体 3; 胆管上皮细胞; 原发性胆汁性肝硬化; 内源性活化

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Role of Endogenous TLR3 Activation in Primary Biliary Cirrhosis

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ABSTRACT Objective: To investigate the effect of Toll like receptor 3 (TLR3) on the injury of primary biliary cirrhosis in bile duct epithelial cells. **Methods:** The human intrahepatic bile duct epithelial cells (HiBEC) were cultured in vitro, and HiBEC samples were prepared by freezing and thawing treatment. Death samples and the live samples were mixed and cultured for 6h to prepare TLR3 endogenous activated samples, labeled as observation group. After treated with nuclease, the necrotic cells and live cells were mixed and cultured for 6h to prepare a control sample, labeled as control group. The apoptosis rate, the expression level of TLR3 and the expression level of β interferon-Toll-like receptor domain adaptor protein (TRIF) and the Caspase-3 activity were detected in the two groups. **Results:** The apoptosis rate of the observation group was significantly higher than that of the control group. The TLR3 and TRIF relative expression of the observation group was also significantly higher than that of the control group. Moreover, the caspase-3 average expression of the observation group was obviously higher than that of the control group. All the above mentioned differences were statistically significant ($P < 0.05$). **Conclusion:** The endogenous activation of TLR3 may lead to HiBEC apoptosis. It may promote the occurrence and development of primary biliary cirrhosis.

Key words: Toll like receptor 3; Bile duct epithelial cells; Primary biliary cirrhosis; Endogenous activation

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前言

HiBEC 细胞的炎性细胞浸润及细胞凋亡是原发性胆汁性肝硬化的早期病理改变之一^[1], 探明此炎性反应的过程, 有助于明确其该病的机制。研究证实 T 细胞、NK 细胞介导的炎性反应能够导致 HiBEC 凋亡^[2], 但上述研究忽视了 HiBEC 直接参与免疫调节的能力, 因此存在一定局限性。报道指出, HiBEC 细胞能够通过自身表达 TLR 及多种粘附分子等途径, 直接参与炎症反应^[3], 以 TLR3 为例, 其能够受坏死细胞释放的 RNA 影响, 出现内源性活化, 可能也参与了原发性胆汁性肝硬化的形

成及发展。本研究采用体外实验, 重点探讨了胆管上皮细胞 TLR3 内源性活化在原发性胆汁性肝硬化中的作用机制。

1 材料与方法

1.1 材料及仪器

主要材料: HiBEC, 由美国 Sciencell 提供; 1640 培养基及胎牛血清, 由美国 Hyclone 提供; 核酸酶, 由美国 Sigma 提供; Caspase-3 抗体, 由美国 BD bioscience 提供; RNA 提取试剂盒, 由美国 Qiagen 提供。主要仪器: FACSCalibur 流式细胞仪, 由美国 BD 提供; PCR 分析仪, 由美国 ABI 提供。

1.2 方法

样本制备: ① 体外培养 HiBEC: 在 37℃、5% CO₂、RPMI 1640 完全培养基条件下培养。取适量 HiBEC, 通过反复冻融制备死亡细胞。按照 2:1 的比例混合死细胞与活体细胞, 4℃ 条件下孵育 6 h 后作为观察组; 按照 2:1 的比例混合死细胞与活体

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细胞,并经核酸酶处理,在4℃条件下孵育6h,作为对照组;同条件下培养36h。

检测指标:①采用流式细胞术检测 HiBEC 凋亡率:收集细胞样本,加入缓冲液(由三羟甲基氨基甲烷 50 mmol/L、NaCl 100 mmol/L、BSA 1%、叠氮化钠 0.02%构成,调整 pH 为 7.4),调节密度至每毫升 50×10^6 个细胞,取 195 μ L 悬液加入 5 μ L Annexin-V,常温静置 10min,再行洗涤,并加入缓冲液 190 μ L,最后加入 20 μ g/mL PI 10 μ L,应用流式细胞仪检测细胞凋亡率,统计早期凋亡率(Annexin-V⁺/PI⁻区域)、晚期凋亡率(Annexin-V⁺/PI⁺区域)及总凋亡率(=早期凋亡率+晚期凋亡率);②采用荧光定量 PCR 技术检测 TLR3 mRNA、TRIF mRNA 水平:收集细胞并提取总 RNA,利用基因库相关数据设计 TLR3 mRNA、TRIF mRNA 序列合成引物,将总 RNA 反转录为 cDNA,应用定量 PCR 扩增,读取阈循环值(Ct),根据 $\Delta Ct = \text{目的基因 Ct} - \text{内参基因 Ct}$ 及 $\Delta \Delta Ct = \text{研究组 } \Delta Ct - \text{对照组 } \Delta Ct$,计

算 $2^{-\Delta \Delta Ct}$ 。③以流式细胞术检测 Caspase-3 活性:计算 Perm/Wash 缓冲液及抗体用量,制备 Perm/Wash 缓冲液,收集细胞后首先以预冷 PBS 缓冲液洗涤 2 次,并调整细胞密度至 $1 \times 10^6/0.5\text{mL}$,冰浴 20 min 后离心弃上清,在室温下以 Perm/Wash 缓冲液洗涤 2 次,最后应用流式细胞仪检测 Caspase-3 活性。

1.3 统计学方法

计量资料按($\bar{x} \pm s$)表示,组间比较采用 t 检验,以 $P < 0.05$ 为差异有统计学意义。

2 结果

2.1 两组 HiBEC 凋亡率对比

观察组早期凋亡率、晚期凋亡率及总凋亡率均明显高于对照组,差异有统计学意义($P < 0.05$)(表 1 与图 1)。

表 1 两组 HiBEC 凋亡率对比($\bar{x} \pm s, \%$)

Table 1 Comparison of the HiBEC apoptosis rate between the two groups($\bar{x} \pm s, \%$)

Groups(n)	early apoptosis rate	later period apoptosis rate	total apoptosis rate
Observation group(3)	15.81 $\pm$ 2.05	18.17 $\pm$ 2.01	33.98 $\pm$ 2.39
Control group(3)	1.13 $\pm$ 0.27	1.75 $\pm$ 0.33	2.88 $\pm$ 0.58
t	12.297	13.962	21.903
P	0.000	0.000	0.000

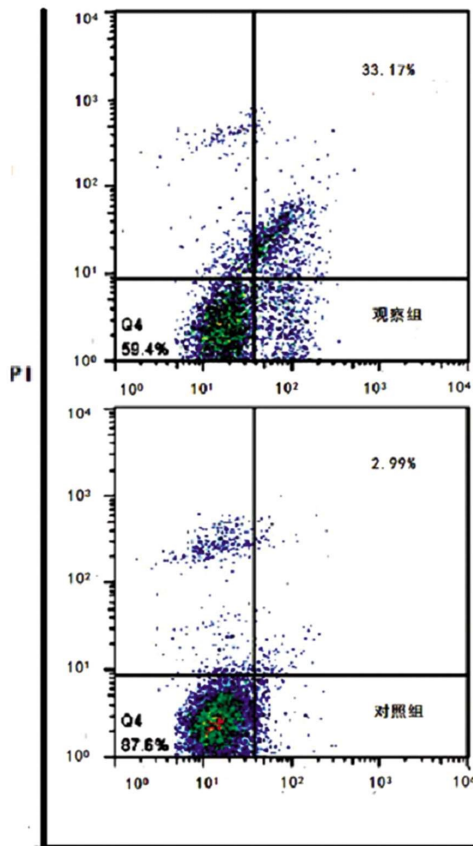


图 1 两组细胞凋亡率对比

Fig. 1 Comparison of the cell apoptosis rate between the two groups

2.2 两组 TLR3 mRNA 及 TRIF mRNA 表达水平对比

观察组 TLR3 mRNA 及 TRIF mRNA 表达水平均明显高于对照组,差异有统计学意义($P < 0.05$)(表 2)。

表 2 两组 TLR3 mRNA 及 TRIF mRNA 表达水平对比($\bar{x} \pm s$)

Table 2 Comparison of the mRNA expression levels of TLR3 and TRIF between two groups($\bar{x} \pm s$)

Groups(n)	TLR3 mRNA	TRIF mRNA
Observation group(3)	3.77 $\pm$ 0.25	4.17 $\pm$ 0.18
Control group(3)	0.93 $\pm$ 0.07	1.05 $\pm$ 0.07
t	18.947	27.981
P	0.0000	0.0000

2.3 两组细胞 Caspase-3 活性对比

观察组 Caspase-3 阳性率(27.7 $\pm$ 3.1)%,明显高于对照组(8.2 $\pm$ 1.8)%,组间差异有统计学意义($t=9.422; P < 0.001$)。

3 讨论

原发性胆汁性肝硬化为常见自身免疫性疾病,此类疾病的直接病因多为自身免疫系统对自身细胞、组织及器官的免疫耐受丧失,使得免疫反应直接对自身造成损伤^[4,5],目前尚未完全探明此类疾病发病机制。临床研究^[6]显示,原发性胆汁性肝硬化的发病,不仅与适应性免疫有关,还与固有免疫有关,固有免疫可能加速了该疾病的发展。

TLR3 在固有免疫中发挥了关键作用,费允云等^[7]报道显示 TLR3 介导的信号通路刺激 IRF-3 产生 1 型干扰素,进而诱导疾病的发生,但此研究主要通过 polyI:C 活化 TLR3 信号通路,并未涉及 TLR3 的内源性活化;赵立宇等^[8-10]报道也显示 TLR3 作用下,NK 细胞对 HiBEC 的杀伤能力有一定上升,但尚未明确 TLR3 与 HiBEC 主动参与炎症反应的关系。以既往研究为

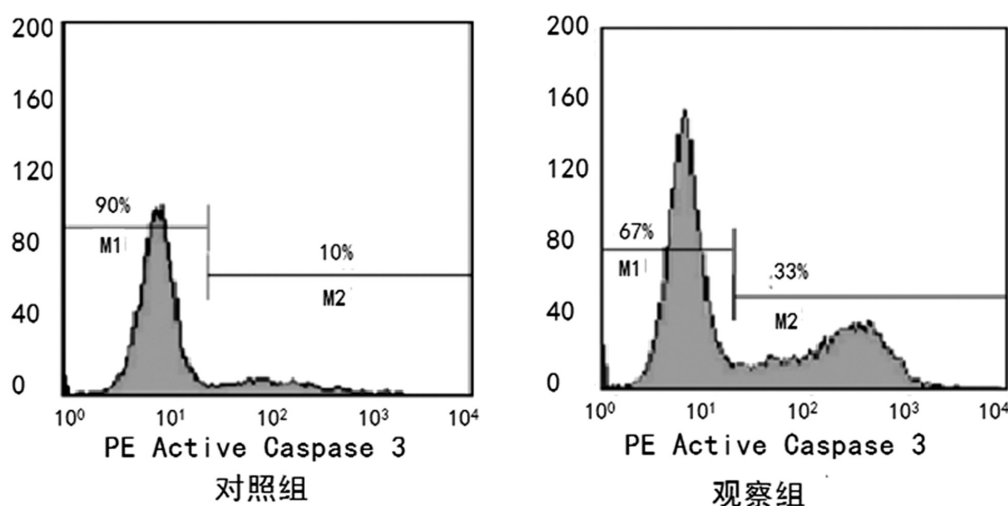


图2 两组细胞 Caspase-3 活性对比

Fig.2 Comparison of the Caspase 3 activity between the two groups

基础,本研究重点分析了TLR3的内源性活化是否与也与HiBEC的炎症反应有关,及该内源性活化的具体机制。

本研究利用坏死HiBEC与活体HiBEC共同培养,制备TLR3内源性活化HiBEC样本,其原理在于HiBEC坏死所释放的内源性RNA能够通过TLR3途径,活化周边正常上皮细胞^[11]。观察组细胞凋亡率明显高于对照组,提示在原发性胆汁性肝硬化早期,少量HiBEC细胞受炎症反应杀伤后,其释放的内源性RNA能够进一步活化周边细胞,导致损伤的迅速扩散,从而加速HiBEC的病理性损伤,上述结果提示,在早期阻断内源性RNA对TLR3途径的活化,可能有助于控制疾病,但具体的阻断方案,可能仍需探讨。

TLR3活化与HiBEC细胞损伤的关系与其信号通路有关^[12-14]。TLR3的信号通路主要有2条,其一为髓样细胞分化因子88依赖性信号通路,该通路下游为丝裂原活化蛋白激酶和Nk- κ B,下游受到激活后,释放TNF- α 及IL-6等细胞因子,可以参与并加剧炎症反应,从而导致HiBEC细胞损伤^[15-17];TLR3还存在一条非髓样细胞分化因子88依赖性信号通路,该通路主要能活化IFN调节因子3,诱导产生IFN- β ,从而直接诱导细胞凋亡,以此损伤HiBEC细胞^[18-20]。本研究观察到观察组TLR3和TRIF mRNA表达量明显升高,提示HiBEC的进展性凋亡过程中,TLR3通路处于激活状态,且由于TRIF主要参与非髓样细胞分化因子88依赖性信号通路,故HiBEC的进展性损伤主要与非髓样细胞分化因子88依赖性信号通路的激活有关,HiBEC的自发性凋亡在原发性胆汁性肝硬化中有一定作用。观察组Caspase-3的表达量上升,则提示TLR3内源性活化所致的细胞凋亡,与Caspase-3途径有关,这有助于明确凋亡发生的信号途径。上述结果提示,阻断非髓样细胞分化因子88依赖性信号通路的激活,可能有助于控制疾病,但本研究并未深入得出具体的阻断方案,可能仍需后续研究补充。

综上所述,本研究证实原发性胆汁性肝硬化早期,HiBEC细胞的局部性损伤可导致内源性RNA的释放,进而活化周边正常细胞TLR3,导致炎症反应的扩大及细胞凋亡率的上升。其中细胞凋亡可能与非髓样细胞分化因子88依赖性信号通路有关,这有助于解释疾病的发生及发展机制。明确上述机制,有助

于为控制疾病提供靶向的干预方式,这还有待后续研究补充。

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