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介绍一种兔 VX2 肿瘤的传代和接种方法及其应用经验和体会*

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摘要 目的:介绍一种兔 VX2 肿瘤的传代和接种方法及其应用经验和体会,从而更好的利用此模型进行生物医学研究。**方法:**从荷瘤新西兰大白兔取活性良好的肿瘤组织块,制备肿瘤细胞悬液,过滤后接种于健康成年新西兰兔左后肢肌肉内。通过一般观察、MRI 和大体及病理学切片对肿瘤进行验证。**结果:**肿瘤传代和接种后生长良好,MRI、大体及组织学验证保持了 VX2 的肿瘤特点。**结论:**本研究介绍的兔 VX2 肿瘤的传代与接种方法稳定、可靠,值得大家推广和应用。

关键词:兔;VX2 肿瘤;接种;模型

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A Method for Passage and Inoculation of VX2 Tumor in Rabbits and Its Application Experiences*

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ABSTRACT Objective: To introduce a method for passage and inoculation of VX2 tumor in rabbits and summarize the experiences with the purpose of its better usage in biomedical research. **Method:** The tumor cell suspensions were prepared from the tumor tissue with excellent viability which dissected from a New Zealand white rabbit bearing the transplantable VX2 carcinoma intramuscularly in the left hind limbs. The general observation, MRI examination, gross and pathological examination were performed to confirm the tumor features. **Result:** The growth of VX2 tumors was excellent after intramuscular passage and inoculation of VX2 tumor cell suspensions in the left hind limbs. Also, MRI finding, gross and pathological observation all confirmed the characteristics of VX2 tumor in this rabbits. **Conclusion:** The method for passage and inoculation of VX2 tumor in rabbits that introduced in this paper proves to be stable and reliable, and it is worthy of wide application in the related field.

Key words: Rabbit; VX2 tumor; Inoculation; Model

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前言

肿瘤动物模型是研究肿瘤发病机制、抗肿瘤治疗和疗效评估研究的重要工具,广泛应用于肿瘤的相关生物医学研究^[1-5],目前常用的肿瘤动物模型包括自发性、诱发性、移植性和转基因动物模型等^[6]。VX2 肿瘤模型是由 Shope 和 Hurst^[7]首次报道的兔乳头状瘤病毒诱发的鳞状细胞癌动物模型,作为大型动物肿瘤模型广泛的应用于肿瘤的发病机制、血管生成、疗效研究、影像学等多种肿瘤生物学行为^[1-5,8-11]。由于 VX2 肿瘤细胞系的建立困难及可能影响肿瘤的生物学特性^[12-16],目前对 VX2 肿瘤的传代多采用组织块传代或细胞悬液注射法^[2,3,11,17,18]。如何长久的传代和接种此模型对于后续研究具有重要意义。本研究

组自 1998 年开始运用 VX2 兔动物模型进行相关研究^[19],现将本研究课题组多年来的传代及接种方法进行介绍,并将经验和体会进行总结以便更好的利用此模型进行更多生物医学研究。

1 材料与方法

1.1 实验动物

成年的体重约 2.0-2.5 kg 健康新西兰大白兔(购自上海宝牧实验动物厂,上海交通大学附属第一人民医院实验动物中心饲养),雌雄均可,适应环境饲养约 1-2 周。

1.2 兔 VX2 肿瘤细胞悬液的制备

取上一代左后肢肌肉 VX2 荷瘤兔(上海交通大学附属第一人民医院放射科保种),瘤体约 2-2.5 cm 直径左右,氯胺酮和

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速眠新 II 复合麻醉剂按 0.4 mL / kg 剂量肌肉注射麻醉。将荷瘤兔仰卧于动物手术台上,局部剃毛后进行常规消毒,铺无菌手术洞巾。然后用手术刀切开皮肤及皮下组织,暴露左后腿肌肉后,止血钳钝性分离肿瘤,用眼科剪取肿瘤包膜下肿瘤边缘呈鱼肉状的新鲜肿瘤组织约 0.5-1 cm³ 大小置于无菌容器内。先用无菌生理盐水冲洗三遍肿瘤组织去除血液,然后用眼科剪不停的将瘤组织尽量剪碎成匀浆状,加入适当生理盐水,置于 200 目无菌不锈钢滤网上去除肿瘤残渣,用无菌 5 mL 注射器吸取过滤后的底层细胞悬液,进行细胞计数和后续的接种,一般细胞悬液的浓度为 10⁶-10⁸ 个 / mL 较为合适。

1.3 兔 VX2 肿瘤的传代接种

取适应饲养 1-2 周后的健康兔 5 只,麻醉及左后肢备皮方法同 1.2,然后将前述调整好的肿瘤细胞悬液约 0.2-0.5 mL(根据实验要求的肿瘤生长速度调整注射量)注射于左后肢肌肉内,拔针后消毒棉签按压针道片刻。待兔清醒后返回饲养笼常规饲养、观察。

1.4 兔 VX2 肿瘤传代后的观察与验证

1.4.1 一般观察

接种后观察动物的活动、状态及饮食情况及肿瘤生长情况。如接种兔出现明显的厌食、活动差或肿瘤增大表面破溃时及时将动物进行安乐死。

1.4.2 MRI 验证

待接种兔左后肢接种处可触及肿瘤并生长至约 2 cm 直径大小时,取荷瘤兔一只,采用氯胺酮和速眠新 II 复合麻醉剂按 0.4 ml / kg 剂量肌肉注射麻醉后,左耳缘静脉置入留置含肝素套管针固定后备用。取仰卧位将荷瘤兔置于 GE HD 3.0 T 磁共振扫描仪检查床上,将双后肢置于膝关节线圈中心,头先进,定位后进行 T1WI、T2WI 和 DWI 序列扫描;然后自左耳缘静脉团注造影剂 Gd-DTPA(马根维显,拜耳先灵药业)按 0.1 mmol/kg 进行 T1WI 增强扫描,图像上传后观察和分析。

1.4.3 大体及组织学验证

取上述 MRI 检查后的荷瘤兔,补充少量麻醉剂深麻下,仰卧位固定,采用 4 % 中性甲醛固定液左心室灌注法^[9]固定后分离肿瘤组织或者直接钝性分离左后肢肿瘤组织后置 4 % 中性甲醛固定液中固定 24 小时以上。然后肿瘤大体标本观察和拍照,沿着肿瘤最长轴切开进行剖面观察和拍照。最后,肿瘤组织常规进行脱水、石蜡包埋和切片,脱蜡后进行常规苏木素-伊红(HE)染色,切片置显微镜下观察细胞形态学特征。

2 结果

2.1 一般观察

接种后接种兔未见明显厌食、嗜睡、活动减少。接种处未见明显红肿及破溃。接种后约 1 周,兔右后肢股后肌群可触及肿瘤。之后随着时间增长,肿瘤逐渐增大。约 2 周左右,肿瘤可长大至 1-2 cm 左右。成瘤率约 95-100 %。

2.2 VX2 荷瘤兔的肿瘤的 MRI 特征

VX2 荷瘤兔 MRI T1WI 图像上肿瘤信号稍高于肌肉信号,有较为清晰的边界(图 1A);T2WI 呈边界较清晰的高信号,中央见小条点状低信号坏死(图 1B);增强后肿瘤较为均匀的明显强化(图 1C),提示肿瘤明显的血供成分;DWI 呈现高信号(图 1D)。

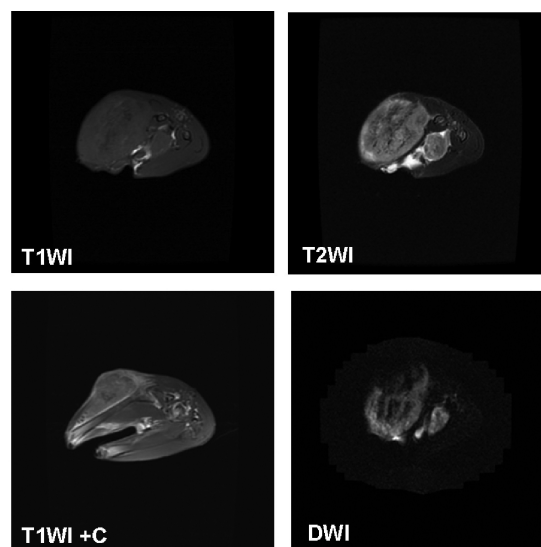


图 1 兔 VX2 肿瘤的 MRI 特征

注:A T1WI, B T2WI, C T1WI 增强, D DWI

Fig. 1 MRI findings of VX2 tumor in rabbit

Note: A T1WI, B T2WI, C T1WI enhancement, D DWI

2.3 VX2 荷瘤兔的肿瘤的大体及组织学特征

解剖后左后肢接种的肿瘤位于肌纤维间隙内,大体标本显示有较清晰的边界和包膜(图 2A),切面肿瘤呈灰白色或灰红色,肿瘤中央可见出血及坏死成分(图 2B),光镜下肿瘤细胞大小不一,核染色加深、核浆比例增加、病理性核分裂像和少许坏死(图 2C)。

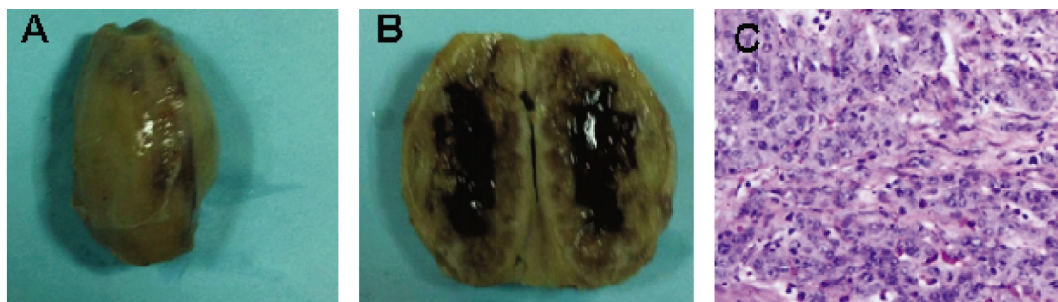


图 2 兔 VX2 肿瘤的大体标本及显微镜切片

注:A 肿瘤整体观, B 肿瘤剖面观, C 光学显微镜切片(HE 染色, × 100)

Fig. 2 Gross and microscopic findings of VX2 tumor in rabbit

Note: A tumor gross sample, B tumor gross section, C microscopic findings(HE staining, × 100)

3 讨论

可靠的肿瘤模型的是保证研究成功的重要前提。本文介绍了一种本实验室十多年来运用的 VX2 肿瘤的传代与接种方法。实践表明,此法传代与接种的肿瘤生长良好。除成功的传代及接种外,本研究组尚应用此法成功建立了 VX2 的血源性脑及脑膜转移瘤模型^[17,20]、脑脊液转移瘤模型^[42]、颌面部转移瘤模型^[1,10,22]等。此方法建立的模型的优点是每次接种的细胞量均一、可控,因此每批次的肿瘤生长速度较均等,形成的肿瘤大小一致,保证了研究基线的一致性。此外,本研究组尚运用组织块接种法建立了 VX2 的肾癌模型^[23]和兔门静脉癌栓形成模型^[23]。本研究组及其他研究表明,VX2 肿瘤可用于肿瘤血管生成^[29]和抗血管生成药物疗效评价^[3]、放疗疗效评估^[1]、影像学研究^[11-5,9,10,17,20-24]等诸多研究领域,是一种较好的大型动物肿瘤研究工具。

在应用本方法时须注意以下事项:① 传代的肿瘤不宜生长过大,一般体积约 2 cm³ 较为合适,如肿瘤已经出现肉眼可见坏死,不宜进行传代;② 选取肿瘤包膜下肿瘤活性充足的肿瘤组织取组织块,取下的组织块要用无菌生理盐水反复冲洗掉血液成分,否则影响肿瘤的生长;③ 在进行肿瘤细胞匀浆的制备时用眼科剪反复剪切且需滤网过滤,但操作时间不宜过长,为保持细胞活性尽量在冰上操作;④ 肿瘤的生长速度与接种量有关,因此可根据研究需要调整细胞的量;如控制肿瘤的生长速度,可将切取的肿瘤组织置于 -20 ℃ 冰箱内 1-2 小时或过夜,可减缓肿瘤的生长速度;⑤ 在进行肿瘤传代与接种时,每批亦 3-5 只为宜,以防止肿瘤丢失,有条件的话每批接种的肿瘤最好进行影像学和组织学验证肿瘤特性。

总之,本文介绍了一种兔 VX2 肿瘤的传代与接种方法及其在本研究组长期以来的应用经验和体会。该方法稳定、可靠,尚可应用于其他的研究模型制备,值得大家推广和应用。

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