

doi: 10.13241/j.cnki.pmb.2020.16.018

## 外周血中性粒细胞 CD4(nCD4)在白血病合并细菌感染患者检测中的应用价值\*

李 娜<sup>1</sup> 张 楠<sup>1</sup> 袁 珍<sup>1</sup> 刘军辉<sup>1</sup> 邱瑞萍<sup>2△</sup>

(1 西安交通大学第一附属医院检验科 陕西 西安 710061;2 西北妇女儿童医院检验科 陕西 西安 710061)

**摘要** 目的:观察外周血中性粒细胞 CD4(neutrophil CD4,nCD4)在白血病合并细菌感染患者检测中的准确度和灵敏度,评价其临床应用价值。方法:筛选我院在2016年1月至2019年1月期间收治的100例白血病患者作为研究对象并纳入研究组,其中合并47例合并细菌感染者纳入研究I组,其余53例未合并细菌感染者纳入研究II组,另外根据血培养检查结果将研究I组分为阳性组( $n=12$ )和阴性组( $n=35$ ),同时,筛选同时期进行健康体检者50例纳入对照组;对比降钙素原(Procalcitonin,PCT)、C-反应蛋白(C-reactive protein,CRP)及nCD4水平,并比较阳性组和阴性组的PCT、CRP、nCD4的诊断准确度和灵敏度。结果:研究组的PCT、CRP、nCD4高于对照组( $P<0.05$ ),研究I组的PCT、CRP、nCD4高于研究II组( $P<0.05$ );研究I组中,阳性组的nCD4荧光强度高于阴性组( $P<0.05$ );nCD4诊断白细胞细菌感染的准确度、灵敏度、特异度优于CRP及PCT诊断。结论:nCD4在白血病合并细菌感染患者检测中具有较好的临床参考价值,联合PCT、CRP检测能够显著提高检测准确性,可以作为诊断白血病合并细菌感染的诊断指标。

**关键词:** 中性粒细胞 CD4(nCD4); 白血病, 细菌感染; 临床检测; 应用价值

中图分类号:R733.7;R446.1 文献标识码:A 文章编号:1673-6273(2020)16-3087-04

## Application Value of Peripheral Blood Neutrophil CD4 (nCD4) in Detection of Patients with Leukemia Complicated by Bacterial Infection\*

LI Na<sup>1</sup>, ZHANG Nan<sup>1</sup>, YUAN Jing<sup>1</sup>, LIU Jun-hui<sup>1</sup>, QIU Rui-ping<sup>2△</sup>

(1 Department of Laboratory Medicine, the First Affiliated Hospital of Xi'an Jiaotong University, Xi'an, Shaanxi, 710061, China;

2 Department of Laboratory Medicine, Northwest Women and Children's Hospital, Xi'an, Shaanxi, 710061, China)

**ABSTRACT Objective:** Observe the accuracy and sensitivity of peripheral blood neutrophil CD4 (nCD4) in the detection of patients with leukemia complicated by bacterial infection, and evaluate its clinical application value. **Methods:** 100 patients with leukemia admitted in our hospital from January 2016 to January 2019 were selected as the research object and included in the study group. Among them, 47 patients with bacterial infection were included in study group I, and the remaining 53 patients without bacterial infection were included. Study II group, according to the results of blood culture examination, the study I components were positive group ( $n=12$ ) and negative group ( $n=35$ ). At the same time, 50 cases of healthy physical examination at the same period were screened into the control group; The levels of PCT, CRP and nCD4 were compared, and the diagnostic accuracy and sensitivity of PCT, CRP, and nCD4 in the positive and negative groups were compared. **Results:** The PCT, CRP, and nCD4 in the study group were higher than those in the control group (all  $P<0.05$ ), and the PCT, CRP, and nCD4 in the study group I were higher than those in the study II group (all  $P<0.05$ ). The intensity was higher than that of the negative group ( $P<0.05$ ). nCD4 is better than CRP and PCT in the accuracy, sensitivity and specificity of leukocyte bacterial infection. **Conclusion:** nCD4 has good accuracy and sensitivity in the detection of patients with leukemia complicated by bacterial infection, and combined with PCT and CRP detection can significantly improve the detection accuracy, which can be used as a diagnostic indicator for the diagnosis of leukemia combined with bacterial infection.

**Key words:** Neutrophil CD4 (nCD4); leukemia, bacterial infection; clinical detection; application value

**Chinese Library Classification(CLC):** R733.7; R446.1 **Document code:** A

**Article ID:** 1673-6273(2020)16-3087-04

### 前言

白血病是造血系统的一种恶性肿瘤,因疾病、化疗、骨髓造血功能下降等因素,白血病患者的免疫功能大幅度降低,对细

\* 基金项目:国家自然科学基金项目(81500219);陕西省科学技术研究发展计划项目(2016SF-217)

作者简介:李娜(1982-),女,硕士研究生,主管检验师,研究方向:临床检验诊断及妇幼健康研究,

电话:13809188968, E-mail:linana521518@163.com

△ 通讯作者:邱瑞萍(1980-),女,本科,中级检验技师,研究方向:临床基础检验,电话:15902933916, E-mail:24878501@qq.com

(收稿日期:2020-02-06 接受日期:2020-02-28)

菌侵入的抵抗地下降,易发生感染,因此患者合并细菌感染风险较高,是导致白血病患者死亡的主要原因,严重威胁患者生命健康<sup>[1,2]</sup>。白血病合并细菌感染无典型症状,发展迅速,且患者普遍预后不佳,因此早期明确诊断、早期积极治疗对于及时治疗和改善预后而言至关重要<sup>[3-5]</sup>。由于临床很难通过患者的临床症状区分感染病原体的不同,目前普遍借助PCT、CRP进行检测,但是上述指标具有与一定的局限性,难以明确诊断细菌感染,诊断价值较低,且会增加细菌耐药性、菌群紊乱的风险<sup>[6,7]</sup>。有文献<sup>[8]</sup>报告nCD4是诊断败血症和细菌感染的可靠指标<sup>[9,10]</sup>。因此,在本研究中,为了能够为临床工作提供指导,我们观察了nCD4在白血病合并细菌感染患者检测中的准确度和灵敏度。现将有关资料整理报告如下。

## 1 临床资料

### 1.1 一般资料

筛选我院在2016年1月至2019年1月期间收治的100例白血病患者为研究组。纳入标准:<sup>①</sup> 初次确诊者,无既往相关病史;<sup>②</sup> 未接受相关治疗;<sup>③</sup> 都同样采用细菌血培养确诊是否存在感染;<sup>④</sup> 患者及家属均知情同意,经医院伦理委员批准。排除标准:<sup>⑤</sup> 已经明确诊断感染病原体(细菌、真菌感染等)者;<sup>⑥</sup> 合并恶性肿瘤者;<sup>⑦</sup> 存在免疫缺陷者;<sup>⑧</sup> 不同意本研究内容者;<sup>⑨</sup> 依从性差者。其中男67例,女33例;年龄24~54岁,平均35.29±5.22岁。其中47例合并细菌感染者纳入研究I组,男22例,女15例;年龄26~54岁,平均36.22±5.27岁;其余53例未合并细菌感染者纳入研究II组,男25例,女28例;年龄24~52岁,平均35.48±5.24岁。另外根据血培养检查结果,将研究I组中12例纳入阳性组,男7例,女5例;年龄27~54岁,平均35.35±5.22岁;35例纳入阴性组,男15例,女10例;年龄25~54岁,平均36.24±5.19岁。另选择同时期进行健康体检者50例纳入对照组,男28例,女22例;年龄25~51岁,平均36.25±5.25岁。各组一般资料比较无差异( $P>0.05$ ),具

有可比性。

### 1.2 研究方法

**1.2.1 检测方法** 两组入院之后常规抽取空腹静脉血。PCT以免疫层析法进行检测,CRP以酶联免疫吸附法进行检测,均应用广州健伦生物科技有限公司提供的试剂盒,nCD4采用上海迪普生物技术有限公司提供的BDFACSAria II流式细胞仪提供,平均荧光强度采用流式细胞术检测,试剂盒采用美国Becton Dickinson公司提供的nCD4流式细胞试剂盒。所有操作严格按照说明书进行。

**1.2.2 指标判定** 对照组检查者的PCT、CRP、nCD4检测数值符合正态分布曲线( $\bar{x}\pm SD$ ),试验组各个亚组的检查结果以高于均值两个标准差( $\geq \bar{x}\pm 2SD$ )及以上者判定为阳性,据此判定上述各个指标在判断白血病合并细菌感染的准确度、灵敏度、特异度。

### 1.3 观察内容

观察不同组的PCT、CRP、nCD4水平以及阳性组和阴性组的准确度、灵敏度、特异度。

### 1.4 统计学方法

应用SPSS 21.0,计量资料以( $\bar{x}\pm s$ ),两组间对比行t检验,多组间行方差分析, $P<0.05$ 为有统计学意义。PCT、CRP、nCD4诊断准确性、灵敏度、特异度比较采用ROC曲线检查,准确度=(真阳性+真阴性)/总例数×100%;灵敏度=真阳性/(真阳性+假阴性)/总例数×100%;特异度=真阴性/(假阳性+真阴性)×100%。

## 2 结果

### 2.1 各组PCT、CRP、nCD4比较

研究组的PCT、CRP、nCD4水平均显著高于对照组( $P<0.05$ ),研究I组的PCT、CRP、nCD4高于研究II组( $P<0.05$ ),见表1。

表1 各组的PCT、CRP、nCD4比较( $\bar{x}\pm s$ )

Table 1 Comparison of PCT, CRP and nCD4 in each group ( $\bar{x}\pm s$ )

Groups	n	PCT(ng/mL)	CRP(mg/L)	nCD4
Study group	I group	47	10.24±1.26*#	72.58±10.68*#
	II group	53	3.29±0.85*	15.22±2.48*
Control group	50	0.45±0.09	2.11±0.12	25.35±6.49

Note: Compared with the control group, \* $P<0.05$ ; compared with the study II group, # $P<0.05$ .

### 2.2 阳性组和阴性组nCD4荧光强度比较

研究I组中,阳性组的nCD4荧光强度为248.65±77.35,显著高于阴性组的55.63±12.29,两组对比差异有统计学意义( $t=9.638, P=0.000, P<0.05$ )。

### 2.3 诊断准确度、灵敏度、特异度比较

经过比较nCD4诊断白细胞细菌感染的准确度、灵敏度和特异度均优于CRP及PCT诊断,见表2。

表2 诊断准确度、灵敏度、特异度比较[例(%)]

Table 2 Comparison of the accuracy, sensitivity, and specificity of diagnosis [n (%)]

Testing indicators	Accuracy	Sensitivity	Specificity
nCD4	91.00(91/100)	95.74(45/47)	86.79(46/53)
CRP	72.00(72/100)	70.21(33/47)	73.58(39/53)
PCT	75.00(75/100)	74.47(35/47)	75.47(40/53)

### 3 讨论

白血病是一种造血干细胞恶性克隆性疾病，会对骨髓的正常造血功能产生抑制，联合化疗是临床治疗该病的重要方法<sup>[11,12]</sup>，但由于缺乏治疗靶向性，在杀死肿瘤细胞的同时，也会杀伤正常血细胞，出现各种并发症，降低患者自身免疫力，极易继发感染，造成系统性炎症反应，严重威胁患者生命健康<sup>[13,14]</sup>。感染是白血病最为棘手的并发症之一<sup>[15]</sup>。联合化疗会导致白血病患者白细胞数量显著降低，造成感染症状不典型，有些合并感染患者只单纯有发热症状，难以早期明确诊断，容易延误治疗时机<sup>[16,17]</sup>。为改善患者预后，早期明确诊断是否合并细菌感染非常必要。

细菌培养、白细胞计数、CRP、PCT 是临床诊断感染性疾病的传统指标，其中，细菌培养是金标准，是当前临床诊断感染性疾病最为客观的指标，但存在一些不足，如检查时间长、某些细菌只有在晚期可以培养出甚至无法培养，另外标本采集时间、部位以及送检时间等因素也一定程度上限制了它的临床应用<sup>[18-20]</sup>。白细胞计数、CRP 是当前临幊上诊断感染性疾病的常用辅助指标，但容易受到其他因素影响，检测的灵敏度普遍不高<sup>[21,22]</sup>。PCT 是较好的感染指示指标，但存在其指示作用存在一定延迟反应，另外，由于白血病患者受输血、集落因子的影响，血清PCT 水平诊断感染情况的准确性、灵敏性、特异性均相对不高<sup>[23,24]</sup>。在本研究中，CRP、PCT 诊断白血病合并细菌感染的准确度、灵敏度、特异度分别为 72.00%、70.21%、73.58% 和 75.00%、74.47%、75.47%，这 2 项指标的准确度、灵敏度、特异度的绝对值也相对较低，证实了上述观点。

近年来的研究发现，中性粒细胞膜表面的 CD4 分子与感染存在密切关系<sup>[25]</sup>。中性粒细胞是机体抵抗感染的主要防御细胞，CD4 分子参与了中性粒细胞的噬菌作用<sup>[26]</sup>。在正常情况下，中性粒细胞表面分子在外周血中呈低水平表达，一旦机体组织发生感染，中性粒细胞被激活，在 4~6 h 会大量表达 CD4。nCD4 与其配体结合之后，会启动并放大免疫效应，在细胞因子的调节下，通过免疫复合物清除作用、细胞吞噬作用、细胞毒性作用来进一步发挥清除病原体、释放介导炎症介质以及抗原递呈的作用<sup>[27-29]</sup>。所以，nCD4 对细菌感染的诊断意义重大。另外，有研究结果<sup>[30]</sup>证实，nCD4 表达与患者的年龄、性别无关，其表达水平高低仅与细菌感染有关，因此，可以适用于新生儿、儿童、成年人不同年龄段以及不同性别的群体。在本研究中，nCD4 诊断白血病合并细菌感染的准确度、灵敏度、特异度分别为 91.00 %、95.74 %、86.79 %，其中准确度、灵敏度不仅均优于 CRP、PCT，其结果的绝对值也相对较高。本研究也存在一定的不足，样本量少，没有对比 nCD4、CRP、PCT 联合诊断价值，在后续研究中需要加大样本量，进一步的深入研究，明确白血病合并细菌感染患者检测中有效的诊断手段，为其诊断提供检测指标。

综上所述，临床研究表明，nCD4 在白血病合并细菌感染患者检测中具有较好的准确度和灵敏度，联合 PCT、CRP 检测能够显著提高检测准确性，可以作为诊断白血病合并细菌感染的敏感的诊断指标。

### 参考文献(References)

- [1] Qu Z, Fang B, Ma G, et al. Significance of procalcitonin in bacterial infections among acute leukemia patients with post-chemotherapy agranulocytosis [J]. Srpski Arhiv Za Celokupno Lekarstvo, 2017, 145 (7-8): 382-386
- [2] Badrul Arefin, Martin Kunc, Robert Krautz, et al. The Immune Phenotype of Three Drosophila Leukemia Models [J]. G3 Genesgenetics, 2017, 7(7): 2139-2149
- [3] Gomez S, Fynn AB, Fernanda S, et al. Early bacterial and fungal infection in children receiving allogeneic stem cell transplantation for acute lymphoblastic leukemia in Argentina [J]. Ped Transplant, 2018, 22(1): e13070
- [4] 吴隼, 马栋, 杨满, 等. 老年急性白血病医院感染的临床特点及危险因素分析[J]. 中华医院感染学杂志, 2014, 24(5): 1174-1175
- [5] Erker C, Huppner AR, Walsh TJ, et al. Successful Treatment of Invasive Conidiobolus Infection During Therapy for Acute Lymphoblastic Leukemia[J]. J Pediatr Hematol Oncol, 2017, 40(7): e446-e449
- [6] 梁利杰, 梁华杰, 孙慧, 等. 急性白血病患者化疗后感染的临床分析[J]. 中华医院感染学杂志, 2014, 24(5): 3744-3746
- [7] Lan Hu, Qiuping Shi, Miao Shi, et al. Diagnostic Value of PCT and CRP for Detecting Serious Bacterial Infections in Patients with Fever of Unknown Origin:A Systematic Review and Meta-analysis[J]. Appl Immunohistochem Mol Morphol, 2017, 25(8): e61-e69
- [8] 张勇刚. T 细胞亚群 CD3、CD4、CD8 和中性粒细胞 CD64 在新生儿细菌感染诊断中的临床意义 [J]. 中国妇幼保健, 2016, 31(9): 1888-1890
- [9] 陈超华. 白血病合并细菌感染患者检测外周血 nCD4 的应用价值 [J]. 临床血液学杂志(输血与检验), 2018, 31(5): 38-40
- [10] Ma SS, Shao LJ, Sun DM, et al. Diagnostic value of neutrophil CD 4 in peripheral blood on leukemia patients with bacterial infection[J]. Chi J Nosocomiol, 2017, 27(4): 777-780
- [11] 骆宜茗, 刘庭波, 谢泗停, 等. 成人急性白血病患者住院化疗感染的临床特征及影响因素研究 [J]. 中华血液学杂志, 2015, 36(12): 1020-1024
- [12] Macanas-Pirard P, Broekhuizen R, González A, et al. Resistance of leukemia cells to cytarabine chemotherapy is mediated by bone marrow stroma, involves cell-surface equilibrative nucleoside transporter-1 removal and correlates with patient outcome [J]. Oncotarget, 2017, 8(14): 23073-23086
- [13] Fellah S, Cheung YT, Scoggins MA, et al. Brain Activity Associated With Attention Deficits Following Chemotherapy for Childhood Acute Lymphoblastic Leukemia[J]. J National Cancer Institute, 2019, 111(2): 201-209
- [14] Krull K, Kunstreich M, Bronsema A, et al. Osteonecrosis in children with acute lymphoblastic leukemia at initial diagnosis and prior to any chemotherapy[J]. Leukemia Lymphoma, 2018, 60(1): 78-84
- [15] Diniz LMO, Maia MMM, Oliveira YV, et al. Study of Complications of Varicella-Zoster Virus Infection in Hospitalized Children at a Reference Hospital for Infectious Disease Treatment [J]. Hospital Pediatrics, 2018, 8(7): 419-425
- [16] Kvaerner AS, Minaguchi J, Yamani NE, et al. DNA damage in blood cells in relation to chemotherapy and nutritional status in colorectal cancer patients-A pilot study[J]. Dna Repair, 2018, 63: 16-24

- [17] Sun W, Triche TJr, Malvar J, et al. A phase 1 study of azacitidine combined with chemotherapy in childhood leukemia: a report from TACL consortium[J]. *Blood*, 2018, 131(10): 1145-1148
- [18] Li C, Yuam BJ, Zhang SQ. Changes of neutrophil CD64 index, C-reactive protein and white blood cell levels in patients with blood bacterial infection [J]. *Chinese J Laboratory Diagnosis*, 2015, 19(10): 1693-1696
- [19] Liat AH, Kfir O, Roy N, et al. A host-protein signature is superior to other biomarkers for differentiating between bacterial and viral disease in patients with respiratory infection and fever without source: a prospective observational study [J]. *Eur J Clin Microbiol Infect Dis*, 2018, 37(7): 1361-1371
- [20] Haruna N, Takao FJ, Suga S, et al. Species differences in circulation and inflammatory responses in children with common respiratory adenovirus infections[J]. *J Med Virol*, 2018, 90(5): 873-880
- [21] Sadettin Er, Büleent Çomçalı, Ahmet Soykurt, et al. Diagnosis of Appendicitis in Patients with a Normal White Blood Cell Count: A Cross-Sectional Study[J]. *Bull Emerg Trauma*, 2018, 6(2): 128-132
- [22] Nelson DA, Hughes JD, Engel CE, et al. Use of Dual-Force Aggregation as a Multiplexed, Rapid Point-of-Care Screening Method for White Blood Cell Counting from Whole Blood Samples[J]. *J Applied Laboratory Med*, 2019, 2(1): 92-97
- [23] Ebihara, Yasuhiro, Kobayashi, et al. Diagnostic performance of procalcitonin, presepsin, and C-reactive protein in patients with hematological malignancies[J]. *J Clin Laborat Analysis*, 2017, 31(6): e22147
- [24] Dai X, Li JP, Li WQ, et al. Changes of Neutrophil CD64 in Patients with Hematological Malignancies Combined with Bacterial Infections[J]. *Zhongguo Shi Yan Xue Za Zhi*, 2017, 25(2): 577-581
- [25] Wang KH, Zang WZ, Li YL, et al. Effect of CD4 (+) T cell surface CD1d molecules on progression of multiple sclerosis in mouse experimental autoimmune encephalomyelitis model [J]. *Zhonghua Yi Xue Za Zhi*, 2018, 98(23): 1873-1875
- [26] Yang F, Feng C, Zhang XD, et al. The Diverse Biological Functions of Neutrophils, Beyond the Defense Against Infections[J]. *Inflammation*, 2016, 40(1): 311-323
- [27] Lv J, Xiong YL, Li WJ, et al. BLT1 Mediates Bleomycin-Induced Lung Fibrosis Independently of Neutrophils and CD4<sup>+</sup>T Cells [J]. *J Immunol*, 2017, 198(4): 1673-1684
- [28] Maria Vono, Ang Lin, Anna Norrby-Teglund, et al. Neutrophils acquire antigen presentation capacity to memory CD4<sup>+</sup>T cells in vitro and ex vivo[J]. *Blood*, 2017, 129(14): 1991-2001
- [29] Elias A Said, Mohammed A Al-Abri, Iman Al-Saidi, et al. Altered Blood Cytokines, CD4 T Cells, NK and Neutrophils in Patients with Obstructive Sleep Apnea[J]. *Immunology Letters*, 2017, 190: 272-278
- [30] Cheng S, Pole JD, Sung L. Early deaths in pediatric acute leukemia: a population-based study [J]. *Leuk Lymphoma*, 2014, 55 (7): 1518-1522

## (上接第 3184 页)

- [15] Varga L, Jovankovicova A, Huckova M, et al. Hereditary bilateral sudden sensorineural hearing loss[J]. *Bratisl Lek Listy*, 2019, 120(9): 699-702
- [16] Fusconi M, Attanasio G, Capitani F, et al. Is there a relation between sudden sensorineural hearing loss and white matter lesions? [J]. *Eur Arch Otorhinolaryngol*, 2019, 276(11): 3043-3049
- [17] Menezes AS, Ribeiro D, Lima A, et al. SCORE risk scale as a prognostic factor after sudden sensorineural hearing loss [J]. *Eur Arch Otorhinolaryngol*, 2019, 276(10): 2739-2745
- [18] Tian G, Zhang S, Yang J. Coexistence of IL-6 -572C/G and ICAM-1 K469E Polymorphisms among Patients with Sudden Sensorineural Hearing Loss[J]. *Tohoku J Exp Med*, 2018, 245(1): 7-12
- [19] Yoon SH, Kim ME, Kim HY, et al. Inflammatory cytokines and mononuclear cells in sudden sensorineural hearing loss[J]. *J Laryngol Otol*, 2019, 133(2): 95-101
- [20] Niu SL, Huang YM, Zhou YQ, et al. Determination of serum HMGB1 and ENA-78 in patients with idiopathic sudden sensorineural hearing loss and its clinical significance [J]. *Chin J of clinicians (Electronic Edition)*, 2011, 5(4): 1059-1062
- [21] Bazzi K, Grierson K, Fagan P. Corticosteroid use in sudden sensorineural hearing loss and the risk of osteonecrosis: a potential medicolegal pitfall[J]. *ANZ J Surg*, 2019, 89(12): 1540-1541
- [22] Bhandari A, Jain S. Early Intratympanic Methylprednisolone in Sudden SNHL: A Frequency-wise Analysis[J]. *Indian J Otolaryngol Head Neck Surg*, 2019, 71(3): 390-395
- [23] Chandrasekhar SS, Hollingsworth DB, Monjur TM, et al. Plain language Summary: Sudden Hearing Loss [J]. *Otolaryngol Head Neck Surg*, 2019, 161(2): 211-217
- [24] Chandrasekhar SS, Tsai Do BS, Schwartz SR, et al. Clinical Practice Guideline: Sudden Hearing Loss (Update) Executive Summary[J]. *Otolaryngol Head Neck Surg*, 2019, 161(2): 195-210
- [25] Sahu ID, Craig AF, Dunagan MM, et al. Probing Structural Dynamics and Topology of the KCNE1 Membrane Protein in Lipid Bilayers via Site-Directed Spin Labeling and Electron Paramagnetic Resonance Spectroscopy[J]. *Biochemistry*, 2015, 54(41): 6402-6412
- [26] Psillas G, Arnaoutoglou M, Gatsios T, et al. Autoimmune recurrent facial palsy and bilateral sudden sensorineural hearing loss following Ramsay Hunt-like syndrome [J]. *Auris Nasus Larynx*, 2012, 39 (2): 229-232
- [27] Inokuchi JI, Go S, Yoshikawa M, et al. Gangliosides and hearing[J]. *Biochim Biophys Acta Gen Subj*, 2017, 1861(10): 2485-2493
- [28] Shen F. Treatment of sudden hearing loss by monosialotetrahexosyl-ganglioside [J]. *J of Otolaryngology and Ophthalmology of Shandong University*, 2013, 27(1): 24-25+30
- [29] Yoshikawa M, Go S, Suzuki S, et al. Ganglioside GM3 is essential for the structural integrity and function of cochlear hair cells[J]. *Hum Mol Genet*, 2015, 24(10): 2796-2807
- [30] Li S, Tan J, Zhang H, et al. Effect of Catgut Implantation on Spatial Learning-memory Ability, Expression of Hippocampal Protein Kinase C Interacting Protein 1 and GluR 2 and Ca<sup>2+</sup> Content in Rats with Chronic Ischemic Cognitive Impairment [J]. *Zhen Ci Yan Jiu*, 2018, 43(6): 347-352