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SDF-1 在乳腺癌患者血清中表达水平及临床意义*

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摘要 目的: 分析 SDF-1 在乳腺癌患者外周血中的表达水平, 探讨其临床意义。**方法:** Elisa 法检测乳腺癌患者术前及术后血清 SDF-1 水平, 分析其与乳腺癌临床病理特征的关系。**结果:** 乳腺癌患者术前(69 例)血清 SDF-1 水平明显高于正常对照组(20 例) (6406.7 ± 1302.5 pg/mL vs 5217.4 ± 1225.7 pg/mL), 有明显统计学差异($P < 0.01$), 发生远处转移的乳腺癌患者(11 例)血清 SDF-1 水平明显高于未发生转移者(58 例) (7656.4 ± 784.1 pg/mL vs 6169.7 ± 1364.6 pg/mL), 差异具有明显统计学意义($P < 0.01$)。ER 及 Her-2 表达阳性乳腺癌的患者 SDF-1 水平较 ER 及 Her-2 表达阴性者低, 差异亦有统计学意义($P < 0.05$)。**结论:** SDF-1 可能是预测乳腺癌发生及远处转移的重要标志物。

关键词: 乳腺癌; Elisa; SDF-1; 远处转移

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Serum Levels of SDF-1 in Breast Cancer and its Clinical Significance*

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ABSTRACT Objective: This study was conducted to investigate the value of the pretreatment circulating stromal cell derived factor-1 (SDF-1) in patients with breast carcinoma. **Methods:** We measured SDF-1 serum concentrations and evaluated the relationship between serum SDF-1 and clinical factors in preoperative and postoperative breast cancer patients. **Results:** Serum levels of SDF-1 is higher in breast cancer patients than in controls ($P < 0.01$), Serum levels of SDF-1 is higher in metastatic patients than non-metastatic patients ($P < 0.01$). Serum levels of SDF-1 is higher in ER and Her-2 positive breast cancer patients than ER and Her-2 negative patients ($P < 0.05$). **Conclusion:** SDF-1 might serve as a possible marker predicting development or distant metastasis in breast cancer.

Key words: Breast Cancer; Elisa; SDF-1; Distant metastasis

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前言

乳腺癌是常见的恶性肿瘤之一, 近年来, 其发病率逐年上升, 已成为女性恶性肿瘤发病率第一位, 虽然其诊断及治疗水平得到了很大提高, 其死亡率较前有所降低, 但是每年死于乳腺癌患者仍较高(5.86/10 万), 术后肿瘤复发及远处转移是导致治疗失败的主要原因。研究表明, 基质细胞衍生因子(stromal cell-derived factor-1, SDF-1)及其特异性受体(chemokine receptor 4, CXCR4)在肿瘤侵袭转移中起重要作用, 其对免疫细胞有趋化作用, SDF-1 表达广泛, SDF-1/CXCR4 生物轴不仅是人体正常生长发育之需要, 也是肿瘤侵袭和转移的重要条件^[1-4]。多项研究表明 SDF-1 在肿瘤的原发及转移灶均有表达, 原发灶 CXCR4 的表达高于转移灶及淋巴结, 而 SDF-1 则相反, 表达 SDF-1 的肿瘤细胞可能在 SDF-1 趋化牵引下, 顺浓度梯度转移至作为配体产生源的某些器官, 形成器官特异性的转移^[1, 4-8]。在对胃癌、结肠癌及骨肉瘤的研究中发现 SDF-1 与肿瘤的生长及

转移关系密切, 且可以预测肿瘤的转移^[9-12]。那么在乳腺癌患者中是否也是如此呢? 因此, 本研究通过检测乳腺癌患者血清中 SDF-1 的表达, 分析其与乳腺癌临床特征之间的关系, 以期为乳腺癌的临床诊疗、预后评估等提供参考依据。

1 材料与方法

1.1 研究对象

选取 2011 年 6 月-2013 年 6 月就诊于上海交通大学医学院附属新华医院(崇明), 经粗针穿刺活检病理确认的乳腺癌者共 69 例为实验组, 其中男 2 例, 女 67 例; 年龄 35~79 岁, 中位年龄 53 岁。按 AJCC-TNM 分期: I 期 4 例, II 期 26 例, III 期 28 例, IV 期 11 例(6 例发生肺转移, 3 例发生肝脏转移, 2 例发生骨转移), 所有实验者就诊前均未采用放化疗。根据乳腺癌患者的肿瘤分期及患者的个人意愿选择(乳腺癌改良根治术, 全乳房切除术), 手术后根据 2011 年版 NCCN 乳腺癌指南进行化疗、放疗、内分泌治疗。20 例健康志愿者为对照组。健康志愿者

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选择标准:年龄与实验者匹配,无重大疾病史、体格检查与常规生化检查均无明显异常。所有参与者均签署志愿书,并经医院伦理委员会审核。

1.2 方法

取乳腺癌患者术前、术后1周与术后三周及对照人群外周血2 mL置于含EDTA抗凝管中,3000转/分4℃离心5分钟,吸取上清液500 μL置于-70℃冰箱至检测。依Elisa操作试剂盒(R&D systems)步骤检测血清中SDF-1水平,具体如下:
a.加样:取待检样品100 μL于已包被之反应孔中,置37℃孵育1小时,洗涤,(同时做空白孔,阴性对照孔及阳性对照孔)。
b.加酶标抗体:于各反应孔中,加入新稀释的酶标抗体(经滴定后的稀释度)100 μL。37℃孵育1小时后洗涤。
c.加底物液显色:加入临时配制的四甲基联苯胺(TMB)底物溶液200 μL于各反应孔中,37℃孵育30分钟。
d.终止反应:分别加入硫酸50 μL于各反应孔中。在(BIO RAD公司,1420 multilabel counter)酶标仪450 nm光源下检测。

1.3 统计学方法

应用SPSS15.0软件行统计学分析,计量数据以 $\bar{x} \pm s$ 表示。两两比较经方差齐性检验后进行单因素方差分析,方差不齐数据采用秩和检验;两组间比较采用t检验,三组及三组以上比较应用Kruskall-Wallis检验,以 $P < 0.05$ 表示差异有统计学意义。

2 结果

血清SDF-1水平:乳腺癌患者($n=69$)(6406.7 ± 1302.5 pg/mL; $2800.5 \sim 9574.6$ pg/mL)明显高于正常对照组($n=20$)(5217.4 ± 1225.7 pg/mL; $4016.9 \sim 6712.1$ pg/mL)($P < 0.01$),发生远处转移乳腺癌患者($n=11$)(7656.4 ± 784.1 pg/mL; $4971.3 \sim 9574.6$ pg/mL)明显高于未发生转移的患者($n=58$)(6169.7 ± 1364.6 pg/mL; $2800.5 \sim 8531.4$ pg/mL)($P < 0.01$),未发生远处转移者亦高于正常对照组($P < 0.05$)(见表1及图1)。

SDF-1的表达与肿瘤分期有显著相关性($P < 0.01$),SDF-1的表达水平与肿瘤分期呈正比,随着肿瘤的分期增加,SDF-1的表达逐渐增加,发生远处转移的患者,SDF-1浓度高达 7656.4 ± 784.1 pg/mL。SDF-1的表达与年龄无明显相关(见表1),SDF-1水平与肿瘤病理分型,肿瘤大小、淋巴结转移亦无明显相关性;而术前及术后三周肿瘤组织表达ER阳性的患者SDF-1浓度显著高于ER阴性的患者,术前(6707.6 ± 1505.7 pg/mL VS 5749.2 ± 1205.4 pg/mL $P=0.021$),术后三周(5951.7 ± 1459.3 pg/mL VS 5025.9 ± 1046.5 pg/mL, $P=0.025$),术后三周Her-2阳性患者的SDF-1浓度显著高于Her-2阴性的患者(6167.7 ± 1146.5 pg/mL VS 5352.4 ± 1359.7 pg/mL $P=0.047$)。术后一周SDF-1浓度及术后三周SDF-1浓度与术前比较均无显著差异性,但有先升后降的抛物线趋势(6365.4 ± 1315.6 pg/mL, 7060.9 ± 1415.5 pg/mL, 5620.1 ± 1342.7 pg/mL)。

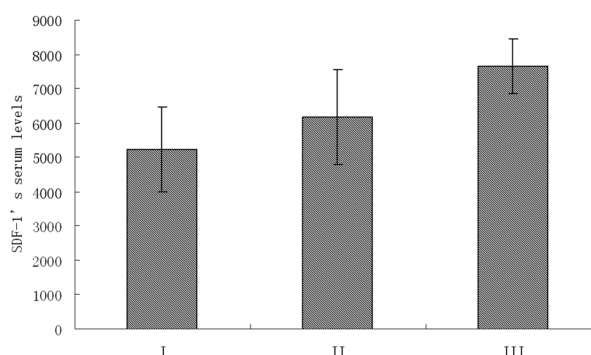


图1 术前各组乳腺癌患者血清中SDF-1表达水平的比较

Fig. 1 Comparison of SDF-1's serum levels of among different group before operation

注: $P < 0.01$, I: 正常对照组; II: 未发生远处转移乳腺癌患者; III: 发生远处转移乳腺癌患者。

Notes: $P < 0.01$, I: Normal control group; II: No distant metastasis in breast cancer patients; III: Distant metastasis in breast cancer patients.

表1 术前乳腺癌患者血清SDF-1浓度与临床特征的关系

Table 1 The relationships between clinical features and SDF-1's serum levels of breast cancer before operation

Charactoristics	n	SDF-1's levels (pg/mL)	P
Age(years)			
≤ 53	35	6626.4± 1284.2	0.207
>53	34	6180.6± 1161.5	
Gender			
Man	2	6728.5± 506.3	6397.1± 1322.7
Woman	67	6397.1± 1322.7	
Stage			
I	4	5318.6± 451.2	< 0.001
II	26	5610.2± 1315.1	
III	28	6810.9± 1127.0	
IV	11	7656.4± 784.1	
Distant metastasis			
present	11	7656.4± 784.1	< 0.001
absent	58	6169.7± 1364.6	
Average concentration	69	6406.7± 1302.5	

表 2 乳腺癌患者手术前后血清 SDF-1 浓度与临床病理特征的关系

Table 2 The relationships between clinical features and SDF-1's serum levels of breast cancer before and after operation

Charactoristics	n	SDF-1's levels		P	SDF-1's levels		P	SDF-1's levels		P
		Preoprerative			Postoprerative(2W)			Postoprerative(3W)		
Age(years)										
≤ 53	36	6626.4±	1284.2	0.184	7129.6±	1426.4	0.802	5627.4±	1284.0	0.873
>53	31	6079.9±	1325.1		6985.8±	1341.6		5612.1±	1318.4	
Tumor diameter										
2 cm	28	6283.6±	1224.3	0.486	7086.4±	1412.8	0.423	5725.3±	1651.3	0.663
>2 cm, ≤ 5 cm	32	6352.7±	1356.8		6908.6±	1607.1		5628.1±	1322.1	
>5 cm	7	6750.7±	941.6		7655.1±	1322.1		5962.7±	1251.7	
Distant metastasis										
M0	9	7626.1±	792.6	0.001	6946.4±	842.7	0.747	5811.7±	781.4	0.566
M1	58	6169.7±	1373.1		7078.7±	1271.4		5590.4±	1374.3	
Cell type										
NIC	8	6388.5±	629.7	0.437	7303.4±	724.1	0.511	5667.8±	761.2	0.895
EIC	53	6307.8±	1151.1		6953.3±	1421.5		5608.1±	1524.5	
OIC	6	6843.4±	860.6		7688.0±	719.6		5662.5±	669.0	
Lymph node metastasis										
N0	46	6574.0±	1208.4	0.224	7264.3±	1344.2	0.176	5468.2±	1246.7	0.251
N1-3	21	5908.3±	1337.2		6615.4±	1328.6		5952.8±	1442.6	
ER										
Present	24	6707.6±	1505.7	0.021	6901.6±	1151.6	0.715	5951.7±	1459.3	0.025
Absent	43	5749.2±	1205.4		7149.8±	1604.8		5025.9±	1046.5	
PR										
Present	35	6641.2±	1085.6	0.180	7152.6±	1427.6	0.684	5733.7±	1506.6	0.428
Absent	32	6063.7±	1475.5		6960.6±	1348.5		5495.9±	1129.2	
Her-2										
Present	22	5852.2±	1286.1	0.061	7201.6±	1451.0	0.691	6167.7±	1146.5	0.047
Absent	45	6616.3±	1415.6		6992.1±	1263.1		5352.4±	1359.7	
Average concentration		6365.4 ± 1315.6			7060.9± 1415.5			5620.1± 1342.7		

NIC=Non-invasive carcinoma, EIC=Early invasive carcinoma, OIC=Other invasive carcinoma.

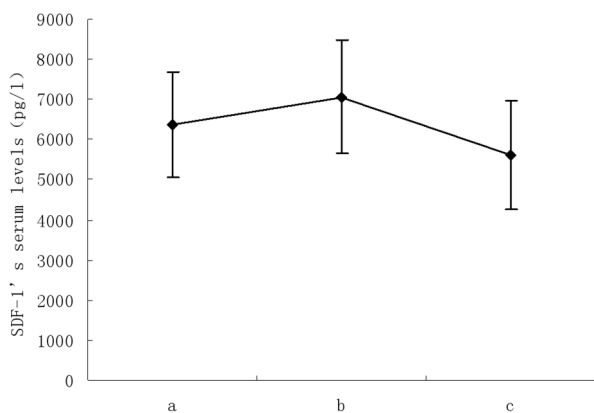


图 2 手术前后乳腺癌患者血清中 SDF-1 表达水平变化

Fig. 2 Changes of SDF-1's serum levels of breast cancer before and after operation

Note: a. Preoperative b. Postoperative after two weeks c. Postoperative after three weeks.

3 讨论

乳腺癌细胞的生长转移与微环境有关,而基质细胞是乳腺癌细胞周围微环境的重要组成部分,乳腺癌细胞生长迅速且易突变,但基质细胞的生长却相对稳定。研究证实,基质细胞分泌的细胞因子可刺激细胞趋化,诱导肿瘤细胞扩散^[1]。

SDF-1 是重要的基质细胞衍生因子,其对免疫细胞有趋化作用,其基因定位于 10 号染色体长臂。SDF-1 表达广泛,在肝、肺、脑、肾、心脏、骨、脑等器官中均有表达,SDF-1/CXCR4 生物轴不仅是人体正常生长发育之需要,也是肿瘤侵袭和转移的重要条件。SDF-1/CXCR4 可通过激活 MEK-MAPK-p42/44, PI-3K-AKT-NF-κ B 等信号通路参与不同的生物学效应^[1-4]。

SDF-1 在结肠癌、胃癌、尤文肉瘤、脑胶质瘤、子宫内膜癌等恶性肿瘤的原发及转移灶亦有表达,原发灶 CXCR4 的表达高于转移灶及淋巴结,而 SDF-1 则相反,表达 SDF-1 的肿瘤细胞可能在 SDF-1 趋化牵引下,顺浓度梯度转移至作为配体产生源的某些器官,形成器官特异性的转移^[1,48]。Woo 等^[9]通过检测胃癌患者血清中 SDF-1 的表达,显示胃癌患者血清中 SDF-1

的表达显著高于正常人,并进一步得出 SDF-1 可作为胃癌患者肿瘤转移的前兆标志物。Nikzaban M^[10] 等研究显示 SDF-1/CXCR4 生物轴的表达与胃癌的分期密切相关。SDF-1 不仅可促进体外结肠癌细胞的转移,还可促进小鼠体内结肠癌转移灶的形成^[11],Krook MA^[12]最新研究发现,在尤文肉瘤细胞中,生长的压力如:无生长因子、低氧及生存空间限制,可以明显提高肿瘤细胞 CXCR4 的表达,后者导致肿瘤细胞沿 SDF-1 浓度梯度向其他脏器转移,以上研究证明 SDF-1 与肿瘤的生长及转移关系密切,且可以预测肿瘤的转移。

那么在乳腺癌患者中是否也是如此呢? Mukherjee D 等^[13] 研究发现 SDF-1 及其受体 CXCR4 在乳腺癌生长及转移过程中起重要作用,通过检测其表达可预测乳腺癌的转移。SDF-1 及其受体 CXCR4 通过促进乳腺癌的粘附,侵袭,乳腺癌细胞随 SDF-1 浓度梯度向靶器官转移^[14]。Müller A 等^[15]在乳腺癌最常见的转移部位淋巴结、肺、肝脏和骨髓中均检测到 CXCR4 高表达,且在 SDF-1 作用下,细胞间丝状肌动蛋白的表达成倍增加,而高表达的肌动蛋白聚合是形成伪足、促进转移所必需的,用抗 CXCR4 的抗体可阻断这些作用。Rhodes LV 等^[16]研究发现 SDF-1/CXCR4 可以显著增加 ER 阳性乳腺癌细胞的雌激素抵抗,并使其恶性程度增加。这些研究说明 SDF-1 在乳腺癌的生长及转移过程中起重要作用。

我们的结果显示,乳腺癌患者血清中 SDF-1 表达显著高于正常人,表明 SDF-1 与乳腺癌之间可能存在着密切相关性;且远处转移者血清 SDF-1 表达水平明显高于未远处转移的患者, SDF-1 的表达随肿瘤的发生及转移逐渐升高,表明 SDF-1 与乳腺癌侵袭转移关系密切;SDF-1 表达随肿瘤分期呈递增趋势,尤其在乳腺癌Ⅲ期及Ⅳ期中表达显著升高,进一步证实 SDF-1 与乳腺癌浸润转移的相关性;乳腺癌术后一周 SDF-1 表达水平较术前有升高趋势,也许与 SDF-1 参与组织修复有关^[1-3],术后三周 SDF-1 水平明显下降,组织修复基本完成, SDF-1 需求减少,表达明显下降,提示乳腺癌手术三周复查 SDF-1 更具临床意义。我们研究结果发现雌激素受体阳性表达的患者 SDF-1 表达较阴性者显著增加,而 Boudot A 等^[17]研究发现,雌激素可以促进 SDF1 及其受体的表达,该研究和我们的结果具有相似性,而且大量研究认为^[18]雌激素水平升高是乳腺癌的危险因素之一,因此雌激素促进了乳腺癌生长及转移,而其中的机制之一可能就是通过促进 SDF-1 的表达,因此我们应用内分泌治疗乳腺癌 ER 表达阳性的患者可以显著抑制肿瘤的复发及转移。HER-2 阳性的乳腺癌患者术后三周 SDF-1 表达显著高于阴性的患者,而 Li YM 等^[19]研究发现在乳腺癌中 HER-2 可以显著提高 SDF-1 的表达,并且认为 HER-2 是通过刺激 SDF-1 的表达促进乳腺癌细胞的生长及转移,术前乳腺癌 HER-2 阳性与阴性之间比较 SDF-1 表达虽然无差异性,但是这可能与样本量少有关。由此可见 SDF-1 在肿瘤的生长及转移中起重要作用,而且 ER 及 HER-2 促进乳腺癌的生长及转移都是通过 SDF-1/CXCR4 信号起作用的,因此应用 SDF-1 或 CXCR4 特异性拮抗剂对治疗乳腺癌患者可能起到意想不到的效果^[20]。

SDF-1 在远处转移的乳腺癌患者中表达更高,提示 SDF-1 可以作为乳腺癌发生转移的重要标志物,这对提高乳腺癌诊断,特别对乳腺癌治疗、手术难度及预后评估,具有明显的临床

使用价值,同时随访乳腺癌术后患者血清中 SDF-1 的表达对评估乳腺癌患者预后也有重要意义。

参考文献(References)

- [1] Domanska UM, Kruizinga RC, Nagengast WB, et al. A review on CXCR4/CXCL12 axis in oncology: No place to hide[J]. Eur J Cancer, 2013, 49(1): 219-230
- [2] Balkwill F. Cancer and the chemokine network [J]. Nat Rev Cancer, 2004, 4(7): 540-550
- [3] Guyon A. CXCL12 chemokine and its receptors as major players in the interactions between immune and nervous systems [J]. Front Cell Neurosci, 2014, 8: 65
- [4] Spano JP, Andre F, Morat L, et al. Chemokine receptor CXCR4 and early-stage non-small cell lung cancer: pattern of expression and correlation with outcome[J]. Ann Oncol, 2004, 15(4): 613-617
- [5] Littlepage LE, Egeblad M, Werb Z. Coevolution of cancer and stromal cellular responses[J]. Cancer Cell, 2005, 7(6): 499-500
- [6] Müller A, Homey B, Soto H, et al. Involvement of chemokine receptors in breast cancer metastasis [J]. Nature, 2001, 410(6824): 50-56
- [7] Walentowicz-Sadlecka M, Sadlecki P, Bodnar M, et al. Stromal derived factor-1 (SDF-1) and its receptors CXCR4 and CXCR7 in endometrial cancer patients[J]. PLoS One, 2014, 9(1): e84629
- [8] Kryczek I, Lange A, Mottram P, et al. CXCL12 and vascular endothelial growth factor synergistically induce neoangiogenesis in human ovarian cancers[J]. Cancer Res, 2005, 65(2): 465-4672
- [9] Woo IS, Hong SH, Byun JH, et al. Circulating stromal cell derived factor-1alpha (SDF-1alpha) is predictive of distant metastasis in gastric carcinoma[J]. Cancer Invest, 2008, 26(3):256-261
- [10] Nikzaban M, Hakhamaneshi MS, Fakhari S, et al. The chemokine receptor CXCR4 is associated with the staging of gastric cancer[J]. Adv Biomed Res, 2014, 3: 16
- [11] Kollmar O, Rupertus K, Scheuer C, et al. Stromal cell-derived factor-1 promotes cell migration and tumor growth of colorectal metastasis[J]. Neoplasia, 2007, 9(10): 862-870
- [12] Krook MA, Nicholls LA, Scannell CA, et al. Stress-induced CXCR4 Promotes Migration and Invasion of Ewing Sarcoma [J]. Mol Cancer Res, 2014 [Epub ahead of print]
- [13] Mukherjee D, Zhao J. The Role of chemokine receptor CXCR4 in breast cancer metastasis[J]. Am J Cancer Res, 2013, 3(1): 46-57
- [14] Claudia Wendel, André Hemping-Bovenkerk, Julia Krasnyanska, et al. CXCR4/CXCL12 Participate in Extravasation of Metastasizing Breast Cancer Cells within the Liver in a Rat Model [J]. PLoS One, 2012, 7(1): e30046
- [15] Müller A, Homey B, Soto H, et al. Involvement of chemokine receptors in breast cancer metastasis [J]. Nature, 2001, 410(6824): 50-56
- [16] Rhodes LV, Bratton MR, Zhu Y, et al. Effects of SDF-1-CXCR4 signaling on microRNA expression and tumorigenesis in estrogen receptor-alpha (ER-α)-positive breast cancer cells[J]. Exp Cell Res, 2011, 317(18): 2573-2581
- [17] Boudot A, Kerdivel G, Habauzit D, et al. Differential estrogen-regulation of CXCL12 chemokine receptors, CXCR4 and CXCR7, contributes to the growth effect of estrogens in breast cancer cells [J]. PLoS One, 2011, 6(6):e20898

- [J]. Chin J Gastroenterol Hepatol, 2010, 19(5): 399-403
- [2] Li M, Li H, Li C, et al. A lpha fetoprotein is a novel protein-binding partner for caspases and blocks the apoptotic signaling pathway in hum anhepatoma cells[J]. Int J Cancer, 2009, 124(12): 2845-2854
- [3] Wang J, Yi Y, Li B, et al. CYFRA21-1 can predict the sensitivity to chemoradiotherapy of non-small-cell lung carcinoma[J]. Biomarkers, 2010, 15(7): 594-601
- [4] Rasch CR, Hauptmann M, Schornagel J, et al. Intra-arterial versus intravenous chemoradiation for advanced head and neck cancer: Results of a randomized phase 3 trial[J]. Cancer, 2010, 116(9): 2159-2165
- [5] Hanagiri T, Sugaya M, Takenaka M, et al. Preoperative CYFRA 21-1 and CEA as prognostic factors in patients with stage I non-small cell lung cancer[J]. Lung Cancer, 2011, 74(1): 112-117
- [6] Chan SL, Mo FK, Johnson PJ, et al. New utility of an old marker: serial alpha-fetoprotein measurement in predicting radio logic response and survival of patients with hepatocellular carcinoma undergoing systemic chemotherapy [J]. JC lin Oncol, 2009, 27(3): 446-452
- [7] Pei GT, Wu CW, Lin WW. Hypoxia induced decoy receptor 2 gene expression is regulated via a hypoxia inducible factor 1 alpha-mediated mechanism[J]. Biochem Biopsy Res Commun, 2010, 391(2): 1274-1279
- [8] Wang J, Zhang N, Li B, et al. Decline of serum CYFRA21-1 during chemoradiotherapy of NSCLC:a probable predictive factor for tumor response[J]. Tumour Biology, 2011, 32(4): 689-695
- [9] Wang C, Q i R, Li N, et al. Notch1 signaling sensitizes tumor necrosis factor-related apoptosis inducing ligand-induced apoptosis in human hepatocellular carcinoma cells by inhibiting Akt/H dm2-mediated p53 degradation and up-regulating p53-dependen tDR5 expression [J]. J Biol Chem, 2009, 284(24): 16183-16190
- [10] Molina R, Aug J M, Bosch X, et al. Usefulness of serum tumor markers, including progastrin-releasing peptide, in patients with lung cancer: correlation with histology[J]. Tumour Biol, 2009, 30(3): 121-129
- [11] Cedrs S, Nuez I, Longo M, et al. Serum tumor markers CEA, CYFRA21-1, and CA125 are associated with worse prognosis in advanced non small cell lung cancer (NSCLC)[J]. Clin Lung Cancer, 2011, 12(3): 172-179
- [12] Grunnet M, Sorensen JB. Carcinoembryonic antigen (CEA) as tumor marker in lung cancer[J]. Lung Cancer, 2012, 76(2): 138-143
- [13] Wang K, Chen Z, Meng Z, et al. Analgesic effect of high intensity focused ultrasound therapy for unresectable pancreatic cancer[J]. Int J Hyperthermia, 2011, 27: 101-107
- [14] 黄培宁, 邓有辉. 微生态制剂对慢性肝衰竭患者肠道菌群、血浆内毒素及细胞因子的影响 [J]. 中华医院感染学杂志, 2011, 21(18): 3798-3800
- Huang Pei-ning, Deng You-hui. Effects of probiotics on intestinal flora plasma endotoxin and cytokines in patients with chronic liver failure[J]. Chin J Nosocomiol, 2011, 21(18): 3798-3800
- [15] 姚维敏, 陈焰, 谢春生, 等. 血必净注射液联合胸腺肽 $\alpha 1$ 治疗慢性肝衰竭的疗效观察[J]. 临床和实验医学杂志, 2012, 11(6): 421-423
- Yao Wei-min, Chen Yan, Xie Chun-sheng, et al. Clinical study on Xuebijing combined with Thymosin $\alpha 1$ in treatment of chronic liver failure [J]. Journal of Clinical and Experimental Medicine, 2011, 11(6): 421-423
- [16] Sung HY, Jung SE, Cho SH, et al. Long-term outcome of high-intensity focused ultrasound in advanced pancreatic cancer [J]. Pancreas, 2011, 40: 1080-1086
- [17] Cui P, Yu M, Peng X, et al. Melatonin prevents human pancreatic carcinoma cell PANC-1-induced human umbilical vein endothelial cell proliferation and migration by inhibiting vascular endothelial growth factor expression[J]. J Pineal Res, 2012, 52: 236-243
- [18] Cho SY, Lee HJ, Jeong SJ, et al. Sphingosine kinase 1 pathway is involved in melatonin-induced HIF-1 α inactivation in hypoxic PC-3 prostate cancer cells[J]. J Pineal Res, 2011, 51: 87-93
- [19] Hyung-Ran Kim, Kyoung-Ho Lee, Su Jung Park, et al. Anti-cancer activity and mechanistic features of a NK cell activating molecule[J]. Cancer Immunol Immunother, 2009, 58(10): 1691-1700
- [20] Cabrera J, Negrí n G, Esté vez F, et al. Melatonin decreases cell proliferation and induces melanogenesis in human melanoma SK-MEL-1 cells[J]. J Pineal Res, 2010, 49: 45-54

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- [18] Chlebowski RT, Anderson GL. Changing concepts: Menopausal hormone therapy and breast cancer [J]. J Natl Cancer Inst, 2012, 104(7): 517-527
- [19] Li YM, Pan Y, Wei Y, et al. Upregulation of CXCR4 is essential for HER2-mediated tumor metastasis [J]. Cancer Cell, 2004, 6(5): 459-469
- [20] Ling X, Spaeth E, Chen Y, et al. The CXCR4 antagonist AMD3465 regulates oncogenic signaling and invasiveness in vitro and prevents breast cancer growth and metastasis in vivo[J]. PLoS One, 2013, 8(3): e58426